

## MedaFluo™ Mounting Medium: MM001M, MM001L

**Intended Use:** For research use only

**Introduction:** MedaFluo™ Mounting Medium is an aqueous based mounting medium for preserving fluorescence staining signal on tissue section and cell smears. It contains anti-fade agent and prevents rapid photo bleaching of fluorophores. It is compatible with Medaysis dyes, DyLight®, FITC, Texas Red, AMCA, Cy® dyes, Alexa Fluor®, fluorescent nuclear stains, fluorescent proteins, fluorescent tracers, and most other fluorophores etc. The fluorescence is retained during prolonged storage at 4°C in the dark. It does not contain phenylenediamine which destroys immunofluorescence of Cy dyes, R-PE, PC and APC.

**Reagent Provided:**

**Reagent Descriptions**

MedaFluo™ Mounting Medium (ready to use)

**MM001M**

1 x 15 ml

**MM001L**

1 x 100 ml

**Storage and Stability:** Store at 2-8°C or freeze at -20°C. Do not use the reagents if the expiration dates on the label have passed. Do not mix the reagents from different lot. If store at 2-8°C the reagent will be good for about 12 months. Freeze at -20°C will last around 24 months.

**Warnings and Precautions:**

1. For professional use.
2. The Material Safety Data Sheet is available upon request.
3. This product contains anti-fading agent and 0.05 % sodium azide as preservative. At this concentration, it is not classified as hazardous. However, NaN<sub>3</sub> may react with lead and copper plumbing to form highly explosive build-ups of metal azides. Upon disposal, flush with large volumes of water to prevent metal azide build-up in plumbing.
4. Wear appropriate Personal Protective Equipment to avoid contact with eyes and skin.
5. Unused solution should be disposed of according to local, state and federal regulations.

**Protocol Recommendations:**

This product is used after immunofluorescence staining, should be applied when the specimen is still wet.

Bring MedaFluo™ Mounting Medium vial to room temperature.

1. Rinse slide to be mounted with DISTILLED OR DEIONIZED WATER, touch the edges of slide on a paper towel to remove excess water. Place slides on a flat surface away from light.
2. Avoid air bubble from the medium. Apply 3-4 drops of mounting medium directly on top of the specimen and spread out evenly by tilting slide back and forth or spread evenly with a pipette tip making sure the tissue is not touched. Excess medium can be removed by touching the edges of slide against paper towel.
3. Let stand at room temperature for about 5 minutes.
4. Apply coverslip carefully avoiding air bubbles.
5. The specimen is ready for visualization under a fluorescent microscope. Be careful the coverslip can move at this point.
6. User can seal the edges of cover slip with nail polish or any organic medium. It is recommended to let slide sit flat in the dark under room temperature overnight before apply sealing media for optimal performance.
7. For storage it is recommended that the slide be stored in the dark at 2-8°C.

**Limitations:**

Immunofluorescence is a multistep process and good results will depend on the proper handling and processing of the tissue both prior to and during staining. Improper fixation, freezing, thawing, washing, drying, heating, sectioning or contamination with other tissues or fluids may produce artifacts, antibody trapping, or false negative results. Inconsistent results may be due to variations in fixation and embedding methods, or to inherent irregularities within the tissue. Our warranty is limited to the actual price paid for the product. We are not liable for any property damage, personnel injury, time, effort or economic loss due to use our product.

**References for reducing autofluorescence:**

1. <http://www.uhnresearch.ca/facilities/wcif/PDF/Autofluorescence.pdf>
2. Herms J. Rommijn et al. Double Immunolabelling of Neuropeptides in the human hypothalamus as analyzed by confocal laser scanning fluorescence microscopy. The Journal of Histochemistry & Cytochemistry. Volume 47(2): 229-235, 1999.